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(54) Title: CHIMERIC DNA-RNA CATALYTIC SEQUENCES

#### DRDRD-1

5' GGUGCGAGAGCGUCAGUAUUAAGCGG 3' - HIV 792-817 3' CCACGCTCTCGCA) TCATAATTCGCC 5'

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#### (57) Abstract

This invention provides chimeric DNA/RNA catalytic molecules useful to cleave RNA sequences. The invention specifically provides two different chimeric DNA-RNA-DNA RNA-DNA catalytic molecules which are targeted to cleave HIV-1 RNA sequences. These chimeric molecules include DNA sequences which flank a catalytic RNA center. Interaction with the HIV-1 substrate RNAs is achieved by Watson-Crick base pairing of the DNA flanking sequences with HIV-1 RNA. The catalytic ribonucleotide center cleaves the phosphodiester bond of the substrate HIV-1 RNA at the expected location.

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#### CHIMERIC DNA-RNA CATALYTIC SEQUENCES

This application is a continuation in part of Application Serial No. 401,613 filed August 31, 1989.

#### Field of the Invention

This invention pertains to DNA-RNA catalytic molecules. More particularly the invention pertains to chimeric DNA-RNA-DNA-RNA-DNA catalytic molecules effective to cleave HIV-1 RNA sequences, for example.

#### Background of the Invention

Ribozymes are structural RNA molecules which mediate a number of RNA self-cleavage reactions. Two distinct trans-acting ribozymes, "hammerhead" and "hairpin," having different secondary structures have been identified. Oncogenes and Aids (1990) [citation] states:

"Another possible synthetic approach is the development of a chimeric molecule containing a ribonucleotide catalytic center and deoxyribonucleotide flanking sequences. It is also conceivable that chimeric catalysts comprised of an RNA catalytic center and DNA flanking sequences will retain biological activity while having greater stability."

Perreault, et al., Nature, 344:565-567 (1990), describes certain mixed deoxyribo and ribooligonucleotides with catalytic activity. No RNA-DNA catalytic molecules of practical therapeutic utility are known.

#### Summary of the Invention

This invention provides chimeric DNA/RNA catalytic molecules useful to cleave RNA sequences. The invention specifically provides two different chimeric DNA-RNA-DNA-RNA-DNA catalytic molecules which are targeted to cleave HIV-1 RNA sequences. These chimeric molecules include DNA sequences which flank a catalytic RNA center. Interaction with the HIV-1 substrate RNAs is achieved by Watson-Crick base pairing of the DNA flanking sequences with HIV-1 RNA. The catalytic ribonucleotide center cleaves the phosphodiester bond of the substrate HIV-1 RNA at the expected location.

#### General Description of the Invention

In general the catalytic molecules of the invention function as hammerhead or hairpin ribozymes. The preferred molecular construct consists of two known RNA catalytic sequences each flanked by a DNA sequence at the respective 3' and 5' termini and coupled by a DNA sequence at the corresponding 5' and 3' termini. These molecules may accordingly be represented by the formulae I and II:

I.  $3' \times - AAAG - Y - AGUAGUC - Z 5'$ 

or

II. 3' X - CAAAG - Y - AGUAGUC - Z 5' in which X, Y and Z are DNA sequences and AAAG, CAAAG and AGUAGUC are catalytic RNA sequences.

The flanking X and Z components may be any DNA sequences that allow base pairing with the substrate RNA at appropriate positions adjacent to the substrate cleavage site. These flanking sequences may be phosphodiester, phosphorothioate, methyl phosphonate, methyl phosphorate or similar moieties.

Y may be any DNA sequence that base pairs <u>inter</u> se in the manner required for catalytic cleavage of

the substrate by the RNA sequences preferably as shown in base paired form in Formula III:

III. 5' C-G 3' A-T G-C G-C A G

The catalytic molecules of this invention can be synthesized in known manner by commercially available DNA synthesizers such as those produced by Applied Biosystems or Milligen. <u>See</u>, e.g., Perreault, et al, supra.

The X and Z sequences may be substituted at the respective 3' and 5' ends with ligands to facilitate cell entry, targeting within the cell and ultimate stability of the catalysts. Such ligands include by way of example but not of limitation: other nuclotides, proteins, carbohydrates, lipids, steroid hormones and cholesterol.

The catalytic molecules of the invention are administered by known and available delivery agents or systems, including, but not limited to, liposomes, defective viral particles, viral capids, and standard DNA/RNA transfective procedures.

#### Description of the Figures

Figure 1 illustrates one catalytic molecule of the invention base paired to an HIV-1 sequence. The RNA portion of the molecule is encircled.

Figure 2 illustrates a second catalytic molecule of the invention base paired to another HIV-1 sequence. The RNA portion of the molecule is encircled.

Figure 3A depicts a ribonuclease A digestion of the catalytic molecule of Figure 1 as compared with an equivalent all DNA molecule. The conditions were 10 units of commercial (Sigma) pancreatic ribonuclease in 2XSSC buffer added to the oligonucleotides which were in 10 microliters of 50 mM Tric-HCl buffer (pH 8.0). The RNAse was incubated with the sample for 10 minutes before the <sup>32</sup>-P end labelled DRDRD or DNA molecules were electrophoresed in a 15% polyacrylamide gel containing 8M urea. The gel was autoradiographed for 10 minutes to get the exposure depicted.

Figure 3B depicts a cleavage reaction involving the catalytic molecule of Figure 1 under conditions described in Chang, et al., <u>Clinical Biotechnology</u>, <u>2</u>:23-31 (1990).

#### EXAMPLE I

The catalytic molecule of Figure 1 was synthesized in known manner utilizing an automated oligonucleotide synthesizer manufactured by Applied Biosystems, Inc.

The result of ribonuclease A digestion of the catalytic molecule is shown by Figure 3A.

The catalytic molecule produced, as described, was used to cleave each of a 610 nuleotide long (S-610) and a 170 nucleotide long HIV-1 gag transcript. In brief, the buffer was 50 mM Tris-HCl, pH 7.5, lmM EDTA, 10mM MgCl<sub>2</sub> at approximately 1 pmole of target, 3 pmole of ribozyme or DNA. The reactions were carried out at 37°C. for 12 hours. The substrate was either a 610 nucleotide long HIV-1 gag containing transcript (S-610) or a 172 nucleotide long HIV-1 gag containing transcript (S-610). The 5' cleavage product is indicated for both.

In Figure 3B the 5' cleavage product is shown for both transcripts. The 3' cleavage product for the 610 target is not visible due to poor reproduction of

the autoradiograph, but is indicated in its position by a 3' P notation. As a negative control, an all DNA oligonucleotide (D) of the same sequence as the DRDRD molecule was incubated with the same substrates under the same conditions with the result that no cleavage was obtained.

Specific cleavage of an HIV-1 5' LTR splice site with a similar catalytic molecule has also been obtained.

#### CLAIMS

- A catalytic molecule capable of cleaving an HIV-1 RNA sequence at a known ribozyme cleavage site said molecule having the formula
  - 3' X AAAG Y AGUAAGUC Z 5'

or

3' X - CAAAG - Y - AGUAAGUC - Z 5'
in which X and Z are DNA sequences that base pair
with an RNA substrate at positions juxtaposed to said
known cleavage site,

AAAG, CAAAG and AGUAGUC are RNA sequences,

Y is a DNA sequence that base pairs <u>inter</u> <u>se</u> in a manner required to permit said RNA sequences to cleave said substrate at said cleavage site.

- 2. The catalytic molecule shown by Figure 1.
- 3. The catalytic molecule shown by Figure 2.
- A catalytic molecule, as defined by Claim 1, in which said RNA sequence is an HIV-1 sequence.
- 5. A catalytic molecule, as defined by Claim 4, in which said HIV-1 sequence is the HIV-1 sequence shown by Figure 1.
- 6. A catalytic molecule, as defined by Claim 4, in which the HIV-1 sequence is the HIV-1 sequence shown by Figure 2.
- 7. A catalytic molecule capable of cleaving an RNA sequence, said molecule having catalytic RNA moieties linked to first and second DNA moieties which base pair with the substrate RNA sequences flanking the cleavage site and interconnected by a third DNA sequence which base pairs <u>inter</u> se to facilitate said cleavage.

FIG. 1 DRDRD-1

GGUGCGAGAGCGUCAGUAUUAAGCGG 3' - HIV 792-817
CCACGCTCTCGCA) TCATAATTCGCC 5' - HIV 792-817

C UG
A
C UG
B = RNA
C G
C C
A
C C
C C
C C
C C
C C
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## FIG. 2 DRDRD #2

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FIG. 3A

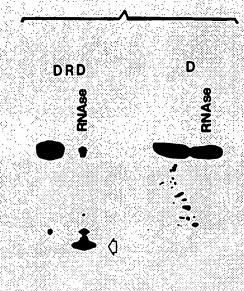
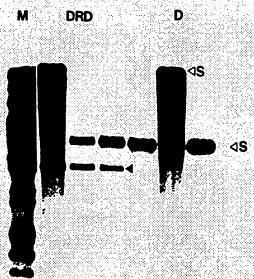


FIG. 3B



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Α	Chemical Abstracts, Volume 110, No. 21, issued 22 May 1989, (Columbus, Chio, U.S.A.) T. R. Cech et al., "RNA Ribozyme Polymerases, Dephosphorylases, Restriction Endoribonucleases and Methods for Their Manufacture", See page 226, column 2, See the abstract No. 187321K, PCT Int. Appl. W08804,300 16 June 1988.	1 - 7
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Category •	Citation of Document, 16 with indication, where appropriate, of the relevant passages 17	Relevant to Claim No
A,P	Nature, volume 344, issued 05 April 1990, J. Peneault, et al., Mixed Deoxyribo - and Ribooligonucleotides with Catalytic activity see pages 565-567.	1-7
A,P	Proceeding of the National Academy of Sciences, Volume 86, no. 23, issued December 1989 (U.S.A.) F.H. Cameron, et al., 'Specific Gene Suppression by Engineered Ribozymes in Monkey Cells', see pages 9139 - 9143.	

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